Study No.: PTGS200219-01 Modified Quantitative Disk Carrier Test Method to Determine the Virucidal Activity of an Ozone-

Generating Whole-Shoe Disinfection Device



STUDY TITLE

Assessment of PathO3Gen Solutions™ Footwear Sanitizing Station for Decontaminating Hard, Non-Porous Environmental Surfaces: Testing against Human Respiratory Coronavirus 229E (ATCC VR-740) as a representative Healthcare-Associated Pathogen

TEST ORGANISM

Coronavirus 229E (ATCC VR-740)

TEST SAMPLE IDENTITY

PathO3Gen Solutions™ Footwear Sanitizing Station

TEST Method

Modified Quantitative Disk Carrier Test Method (ASTM 2197) to Determine the Virucidal Activity of an Ozone-Generating Whole-Shoe Disinfection Device

AUTHOR

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STUDY COMPLETION DATE

March 20, 2020

PERFORMING LABORATORY

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SPONSOR

PathO3Gen Solutions™

STUDY NUMBER

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STUDY PERSONNEL

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TEST SYSTEM

1. Test Microorganism

Coronavirus 229E (ATCC VR-740): Coronavirus 229E (ATCC VR-740) is an enveloped virus in the genus Coronavirus. Members of this genus can cause acute respiratory infections such as SARS-1 and SARS-2 (19-nCOV). Unlike Coronavirus 229E, SARS-1, SARS-2 and Middle-East Respiratory Syndrome (MERS) virus require Biosafety Level 3 labs. Therefore, Coronavirus 229E is frequently used as surrogate for them to assess the activity of different technologies for infection prevention and control (IPAC).

2. Host Cell Line

L-132 cells were used as hosts to support the replication and quantitation of 229E.

The cells were seeded into 12-well multi-well cell culture plates containing modified Eagle's medium (MEM) supplemented with 10% fetal bovine serum (FBS) and maintained at 36±1°C in a humidified atmosphere of 5% CO₂. Efficacy test was performed when the cell monolayer reached >90% confluency.

3. Preparation of Test Inocula

To prepare the virus for inoculation, the virus stock was mixed directly with the soil load (5% FBS). Dilution of the mixture was prepared using Earle's balanced salt solution (EBSS; pH 7.2-7.4).

TEST METHOD

1. Preparation of Test Substance

The efficacy tests were performed on PathO3Gen Solutions[™] Footwear Sanitizing Station following the instruction in the device's user manual at three exposure times (6, 8 and 10 seconds).

2. Test Procedure

A quantitative test system to closely simulate the field-application of the environmental surface decontamination process (modified quantitative carrier test – Tier 2 or QCT-2 (ASTM 2197)) was applied. The protocol was adapted to test the UV LED-based technology. Disks (2 cm diameter) from croc sole shoe were used as archetypical environmental surfaces. Sterile disks were placed on a small platform which was the same size of a shoe at three different positions (middle, back and front). The platform was taped at the bottom of the shoe. The platforms with the disks were exposed to the UV without touching the glass cover of the device. The disks were retrieved in an eluent/neutralizer immediately at the end of the exposure time. The disks were then eluted and the eluates assayed for viable virus.

Each disk on the platform was contaminated with 20 μ L of the virus inoculum with a soil load (5% FBS) and left to dry (contaminated platform) under an operating biosafety cabinet (BSC) for 30±10 minutes. Three disks were contaminated and used as controls.

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Experimental Design

a) Input

The stock virus utilized in the testing was titrated by 10-fold serial dilutions and plaque assayed for infectivity to determine the starting titer of the virus. The results of this control were for informational purposes only.

b) Cytotoxicity Control

Prior to the test, cytotoxicity control and control for interference with virus infectivity were performed to determine if the shoe material caused any apparent degeneration (cytotoxicity) of the host cell line. Control monolayers received an equivalent volume of EBSS (without any neutralizer) only.

c) Efficacy Test

- 1. Disks (2 cm diameter) from croc sole shoes were used in testing of this method, 3 disks were assessed as control without exposure to UV.
- 2. Disks were left inside an operating BSC to dry.
- 3. Disks were inserted on a platform with the same size of the shoe at three different locations (front, middle and back).
- 4. The platform was taped to the bottom of a shoe.
- 5. The experimenter put on the shoes with the platforms at their bottom.
- 6. The experimenter stepped on the device which was already on for 10 minutes.
- 7. After the specific exposure time, the experimenters stepped out of the device.
- 8. The disks were removed from the platform and each disk was placed into a Nalgene vial containing 2 mL of an eluent.
- 9. The L-132 cells in multi-well culture plates were inoculated with 100 μ L of the dilutions prepared from test and control samples. Uninfected indicator cell cultures (cell controls) were inoculated with 100 μ L EBSS alone. The cultures were incubated at 33±1°C in a humidified atmosphere of 5% CO₂ for 40-44 hrs before fixing and staining them for counting the plaque-forming units (PFU).
- 10. Three control disks were included in each test to estimate the initial contamination on the platform. The test was initiated with processing one control before the processing test carriers, one in the middle of the test and ended up with the third control. This was done to take into the account the changes in the input level of the test organisms during the experiment.

DATA ANALYSIS

Calculation of Log₁₀ Reduction

Log₁₀ Reduction = Log₁₀ of average PFU from control carriers – log₁₀ of average PFU the test carriers.

STUDY ACCEPTANCE CRITERIA

No product acceptance criterion was specified for this range-finding study.

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TEST RESULTS

The initial challenge on each carrier were 3.68, 3.73 and 3.65 \log_{10} PFU in three different tests performed on PathO3Gen SolutionsTM Footwear Sanitizing Station. Table 1 show the result of \log_{10} reduction for each contact time. In this test, the drying time of inoculated disks was reduced to 1 hr. In all contact times the \log_{10} reduction was more than 3. No plaque was detected for 8 and 10 seconds contact times.

Table 1: Virucidal Activity Test of PathO3Gen Solutions™ Footwear Sanitizing Station against Coronavirus 229E (ATCC VR-740) with three different contact times

Contact times	Log₁₀ Reduction in PFU								
	Test #1	Test #2	Test #3	Average of Three tests					
6 seconds	3.07	3.28	3.42	3.27					
8 seconds	3.68	3.73	3.65	3.69					
10 seconds	3.68	3.73	3.65	3.69					

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APPENDIX

Result of efficacy test on the device with three different contact times (6, 8 and 10 seconds) against Coronavirus 229E dried on carriers representing shoe soles.

Test #1												
Contac t Time	6 seconds			8 seconds			10 seconds			Control		
Dilution	Front	Middle	Back	Front	Middle	Back	Front	Middle	Back	C1	C2	C3
10 ⁻⁰	0,1,0	1,1,0	0,1,1	0,0,0	0,0,0	0,0,0	0,0,0	0,0,0	0,0,0	TNTC	TNTC	TNTC
10 ⁻¹	0,0,0	0,0,0	0,0,0	0,0,0	0,0,0	0,0,0	0,0,0	0,0,0	0,0,0	21,16,25	25,24, 25	11,16, 23
10 ⁻²	0,0,0	0,0,0	0,0,0	0,0,0	0,0,0	0,0,0	0,0,0	0,0,0	0,0,0	3,3,3	2,2,2	2,2,2
10 ⁻³	0,0,0	0,0,0	0,0,0	0,0,0	0,0,0	0,0,0	0,0,0	0,0,0	0,0,0	0,0,0	0,0,0	0,0,0
10 ⁻⁴	0,0,0	0,0,0	0,0,0	0,0,0	0,0,0	0,0,0	0,0,0	0,0,0	0,0,0	0,0,0	0,0,0	0,0,0

C= Control TNTC= Too numerous to count

Test #2												
Contact Time	6 seconds			8 seconds			10 seconds			Control		
Dilution	Front	Middle	Back	Front	Middle	Back	Front	Middl e	Back	C1	C2	С3
10 ⁻⁰	0,1,1	1,0,0	1,0,0	0,0,0	0,0,0	0,0,0	0,0,0	0,0,0	0,0,0	TNTC	TNTC	TNTC
10 ⁻¹	0,0,0	0,0,0	0,0,0	0,0,0	0,0,0	0,0,0	0,0,0	0,0,0	0,0,0	24,25,25	21,22,24	24,20,25
10 ⁻²	0,0,0	0,0,0	0,0,0	0,0,0	0,0,0	0,0,0	0,0,0	0,0,0	0,0,0	4,3,2	3,2,4	3,3,4
10 ⁻³	0,0,0	0,0,0	0,0,0	0,0,0	0,0,0	0,0,0	0,0,0	0,0,0	0,0,0	0,0,0	0,0,0	0,0,0
10 ⁻⁴	0,0,0	0,0,0	0,0,0	0,0,0	0,0,0	0,0,0	0,0,0	0,0,0	0,0,0	0,0,0	0,0,0	0,0,0
	Test #3											
Contact Time	6 seconds			8 seconds			10 seconds			Control		
Dilution	Front	Middle	Back	Front	Middle	Back	Front	Middl e	Back	C1	C2	C3
10-0	0,0,0	0,1,0	0,1,0	0,0,0	0,0,0	0,0,0	0,0,0	0,0,0	0,0,0	TNTC	TNTC	TNTC
10 ⁻¹	0,0,0	0,0,0	0,0,0	0,0,0	0,0,0	0,0,0	0,0,0	0,0,0	0,0,0	20,22,22	25,25,17	21,19,19
10 ⁻²	0,0,0	0,0,0	0,0,0	0,0,0	0,0,0	0,0,0	0,0,0	0,0,0	0,0,0	2,2,2	2,2,4	3,2,2
10 ⁻³	0,0,0	0,0,0	0,0,0	0,0,0	0,0,0	0,0,0	0,0,0	0,0,0	0,0,0	0,0,0	0,0,0	0,0,0
10 ⁻⁴	0,0,0	0,0,0	0,0,0	0,0,0	0,0,0	0,0,0	0,0,0	0,0,0	0,0,0	0,0,0	0,0,0	0,0,0

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